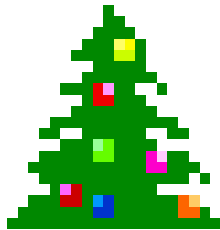


# Molecular Hematopathology

## Lymphomas



December 21, 2004

# Translocations

Small or large fragment of a chromosome fuses with another chromosome

The fusion is viable

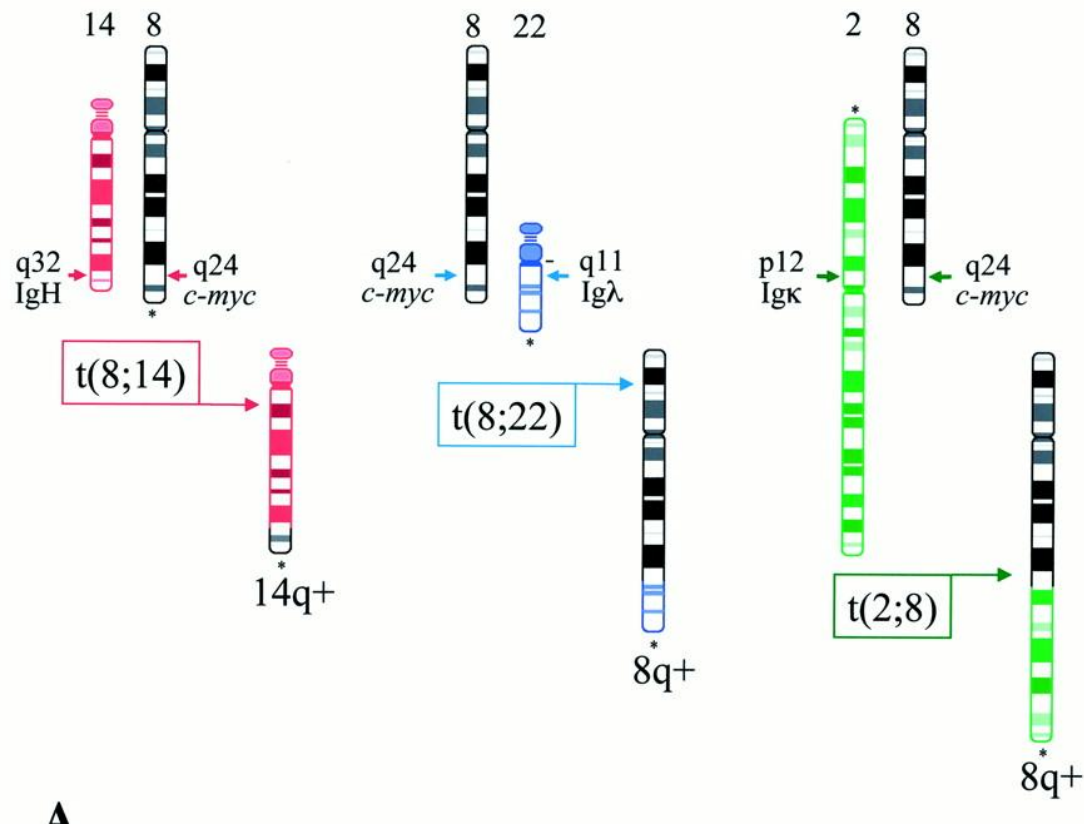
The fusion chromosome is faithfully replicated during subsequent cell generations and, if this gives the cell a growth advantage, a unique large monoclonal population is created

# Translocations

There are 2 general types of translocations seen in non-Hodgkin's lymphomas:

- 1) Intact oncogene on one chromosome translocated beside a gene on another chromosome, usually on an antigen receptor gene, eg Burkitt's or follicular lymphoma
- 2) Two genes on separate chromosomes are disrupted, with portions of both genes fused into one "fusion gene" which encodes a novel protein, eg anaplastic large cell lymphoma

# Detection of *c-myc*/Ig chromosomal translocations in Burkitt's



Hecht, J. L. et al. J Clin Oncol; 18:3707-3721 2000

# Mechanism of Chromosomal Translocation

Mistakes made during B lymphocyte processing

- 1) Marrow - VDJ rearrangement in precursor B cells
- 2) Lymph node - V segment somatic hypermutation
  - IgH class switching
  - receptor editing

Mistakes made during T lymphocyte processing

- 1) Thymus - VDJ rearrangements in progenitor T cells
  - no class switching
  - only rare somatic hypermutation

# B-cell Processing

In order to generate a unique surface Ig, precursor B cells must conjoin V, D, and J segments of genomic DNA on chromosome 14q for IgH and the V and J segments for  $\kappa$  and  $\lambda$  light chains on chromosomes 2p and 22q

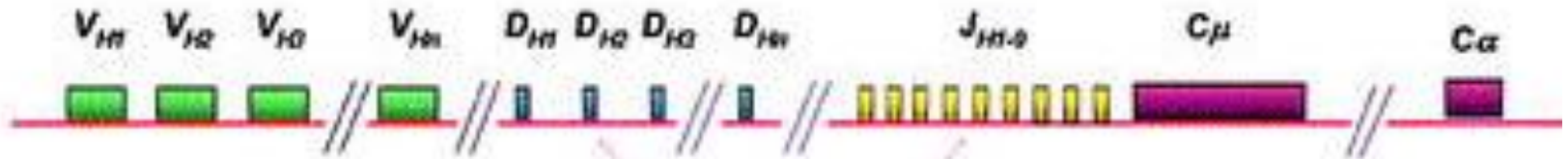
Genomic dsDNA is broken and rejoined by 2 endonucleases (*recombination-activating genes*, RAG 1 and RAG 2) over long distances

dsDNA breaks occur at recognition signal sequences (RSS) that are adjacent to V, D, J sequences

(3' to V, 5' and 3' to D, and 5' to J)

# Chromosome 14q32 of B-cell precursors

## IgH Germline



## D<sub>H</sub>-J<sub>H</sub> Joining

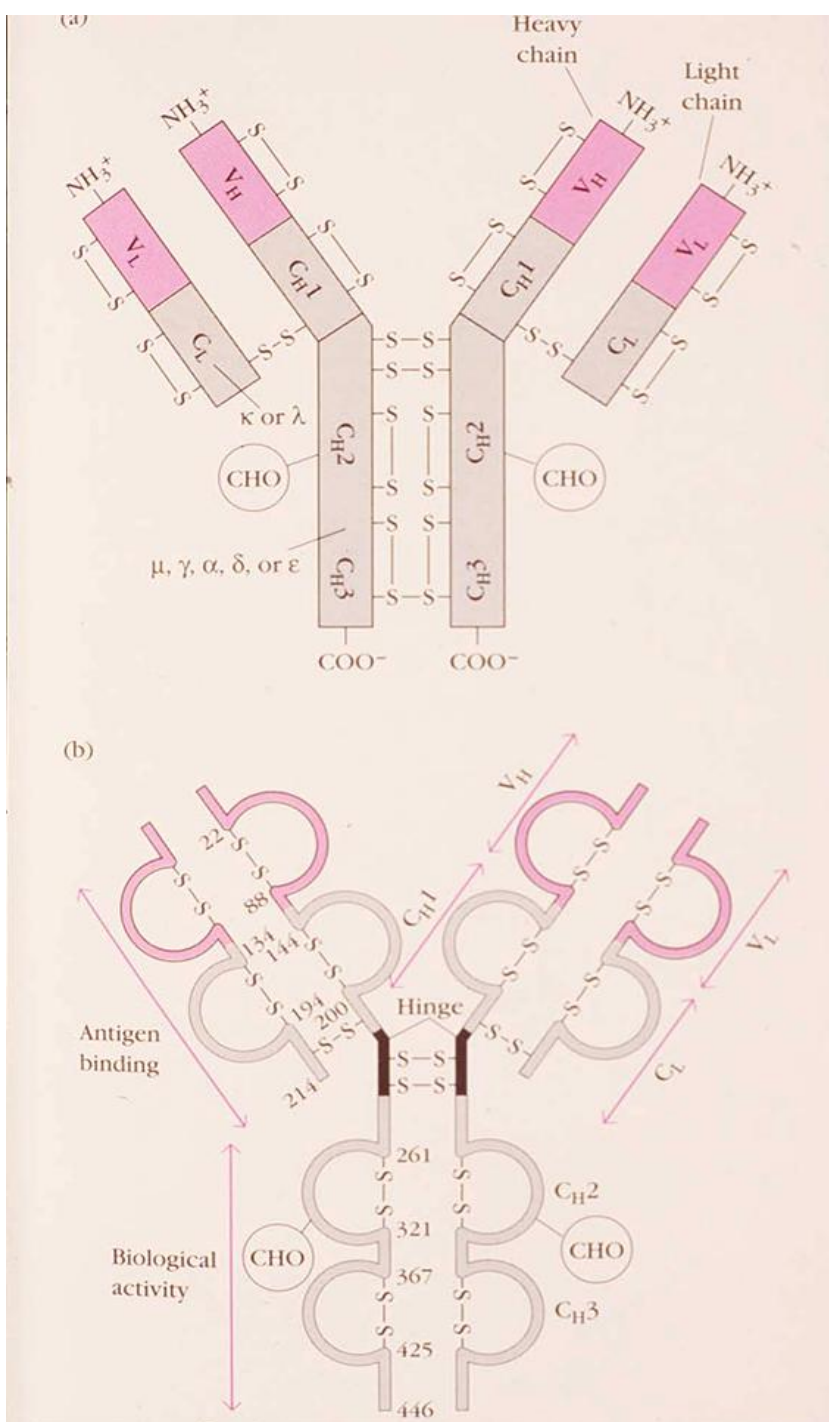


## V<sub>H</sub>-D<sub>H</sub>J<sub>H</sub> Joining



V ~120  
 D ~30  
 J 6  
 C 9

# Immunoglobulin Protein Structure





# Recombination

RSS consists of a 12-13bp spacer region flanked by a conserved 7mer on one end and a conserved 9mer on the other end

Recombination requires one RSS of each type

Recombination also requires:

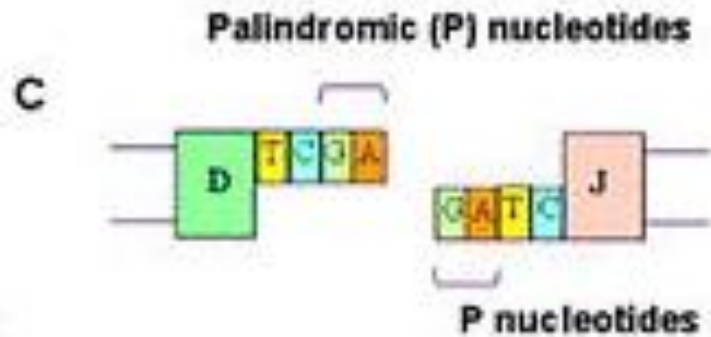
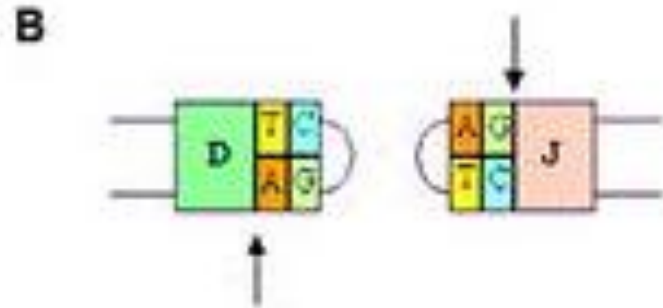
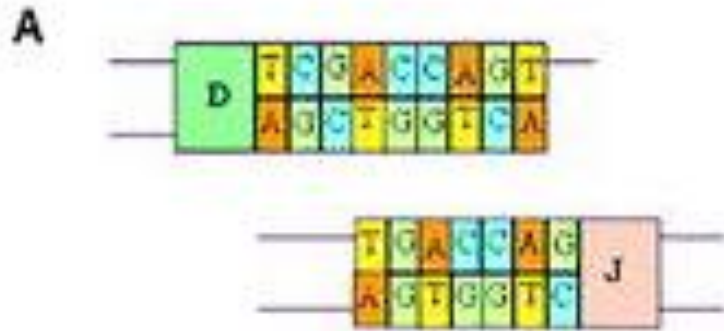
DNA ligase IV

DNA dependent protein kinase

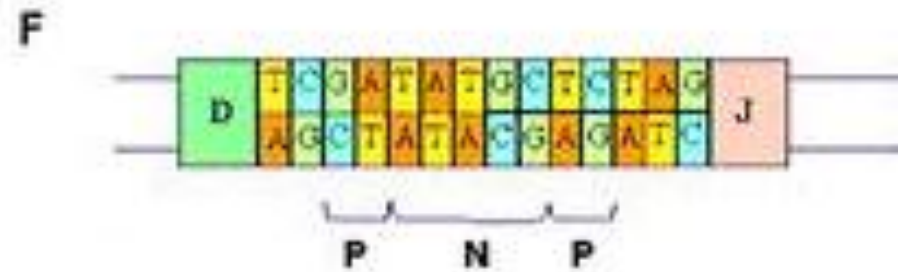
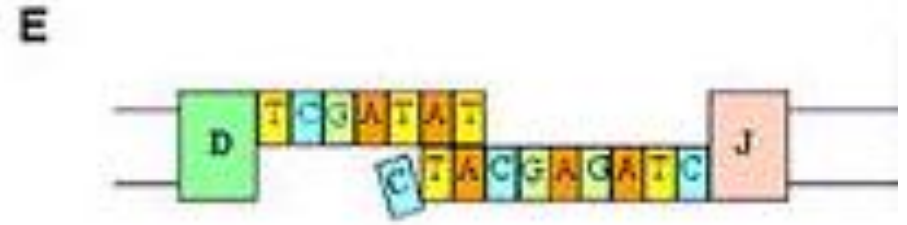
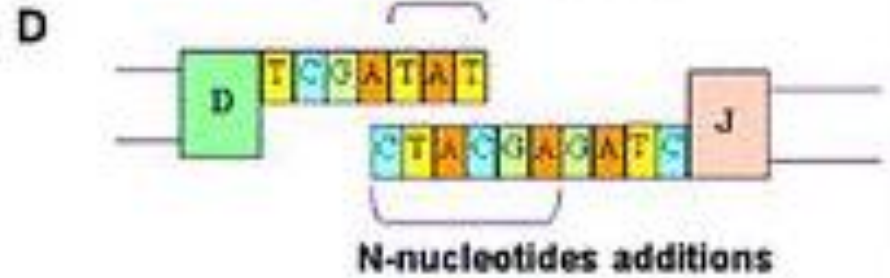
Ku (Ku70/ Ku80 heterodimer), that assoc with DNADPK

XRCC4 (X-ray repair complementary defective in Chinese hamster 4), a DNA repair protein

# Chromosome 14q32 of B-cell precursors



N-nucleotides additions



# VDJ Recombination Steps

RAG breaks the dsDNA and then forms an internal hairpin loop at each end

Hairpin opens up as ssDNA with a 3' overhang

Nucleotides are then added (by TdT) or removed (DNA exonuclease) from the 3' overhang to create diversity

3' overhangs from the two strands then join as the best fit possible to form dsDNA, with some of the nucleotides cut out; AKA “coding end processing”

Gaps filled with DNA polymerase and DNA ligase

# VDJ Recombination Steps

During normal everyday VDJ recombination in these precursor B or T cells, these dsDNA breaks occur

Rarely the dsDNA breaks fuse with the wrong gene → chromosomal translocation

If this translocation puts an Ig or TCR gene next to a proto-oncogene, the cells may acquire a growth advantage and become clonal, eg DLBCL, MCL, BL, precursor T- cell lymphoblastic lymphoma

# VDJ Recombination Steps

At least 2 mechanisms can lead to a translocation:

- 1) Oncogene has a functional RSS sequence that is recognized by RAG, eg in:

precursor T cell lymphoblastic lymphoma  
**t(10;14)(q24;q11)**

- 2) Oncogene gets a dsDNA break by other mechanism of cleavage (not by RAG), eg in:

mantle cell lymphoma  
**t(11;14)(q13;q32)**

# Translocations

Why do they occur in proto-oncogene regions?

Unknown, but theories:

(1) Some oncogenes may have adjacent crossover hotspot instigator sequences that promote recombination, eg: chi sequences around

*bcl-2* of **t(14;18)** follicular lymphoma

*bcl-1* of **t(11;14)** mantle cell lymphoma

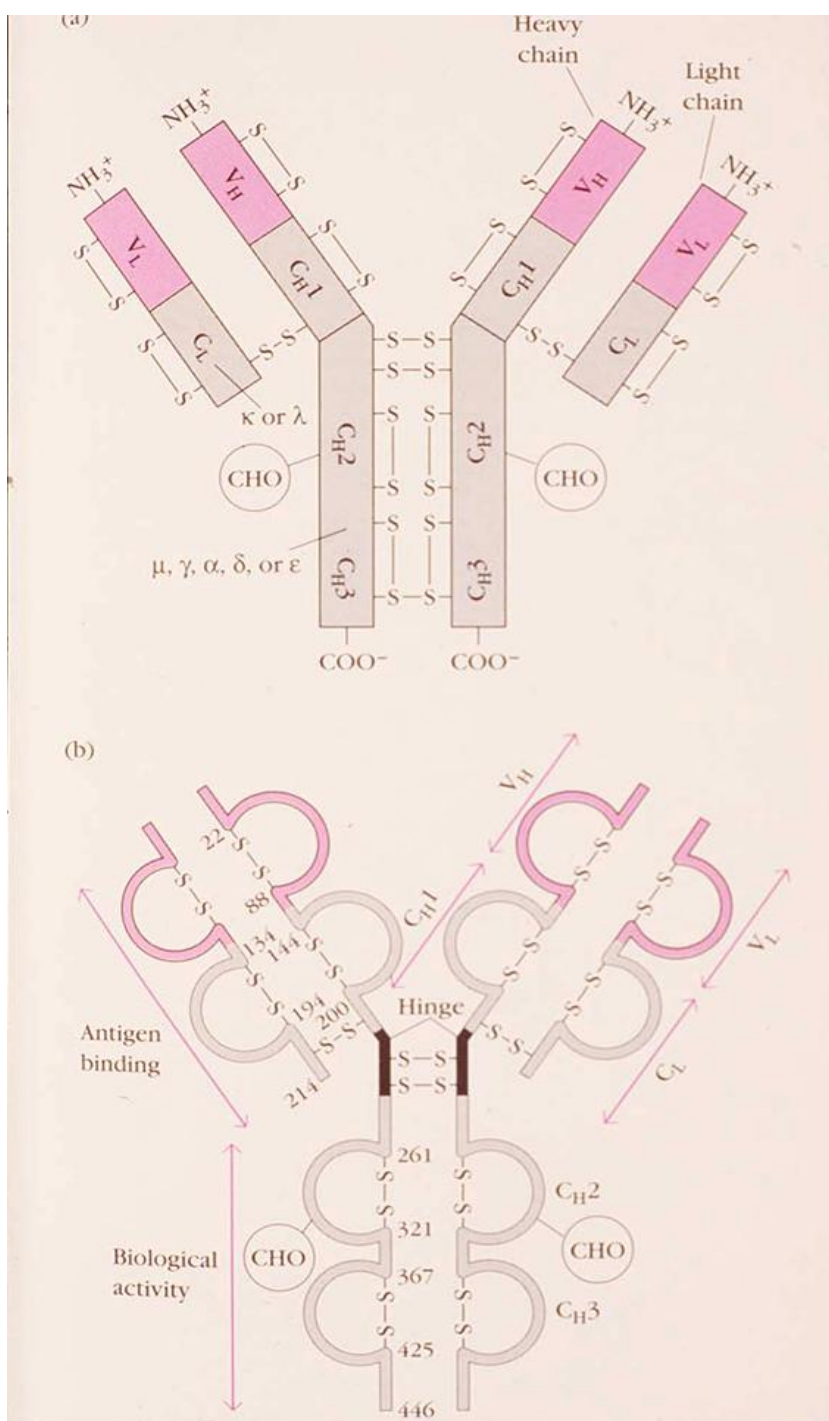
(2) RAG enzyme transpositional activity

# dsDNA Breaks in More Mature B cells

Germinal center cells normally undergo somatic hypermutation

- single nucleotides or short oligos inserted/deleted from V segment by enzyme, after Ag exposure (creates diversity)
- requires dsDNA break
- possibly *c-myc* translocation to IgH gene in endem Burkitt's
- possibly *bcl-6* translocation to IgH gene in DLBCL
- possibly *bcl-2* translocation to IgH in follicular lymphoma

# Immunoglobulin Protein Structure





# dsDNA Breaks in More Mature B cells

Germinal center B cells switch IgM → IgG or IgA or IgE  
after antigen exposure

This recombination requires a dsDNA break in the germ ctr  
DNA between C $\mu$  and C $\gamma$  or C $\alpha$  or C $\epsilon$  is cut out

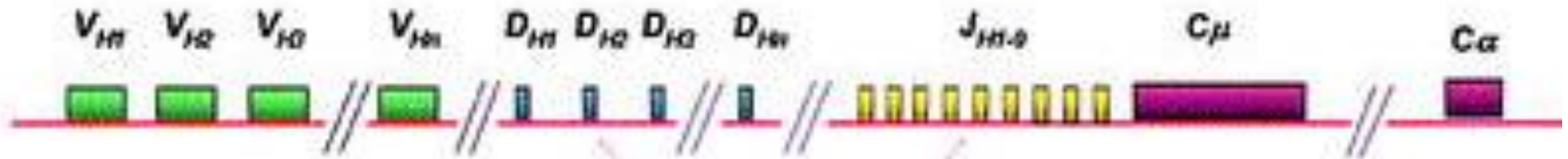
- switch (S) regions around C genes contain microsatellites

Translocations into S regions of IgH described in:

Sporadic Burkitts'	<i>c-myc</i>	<b>t(8;14)(q24;q32)</b>
B-CLL	<i>bcl-3</i>	<b>t(14;19)(q32;q13)</b>
DLBCL	<i>bcl-6</i>	<b>t(3;14)(q27;q32)</b>
PC myeloma	<i>mum/irf4</i>	<b>t(6;14)(p25;q32)</b>

# Chromosome 14q32 of B-cell precursors

*IgH Germline*



*D<sub>H</sub>-J<sub>H</sub> Joining*



*V<sub>H</sub>-D<sub>H</sub>J<sub>H</sub> Joining*



V ~120  
D ~30  
J 6  
C 9

# dsDNA Breaks in More Mature B cells

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DLBCL	<i>bcl-6</i>	<b>t(3;14)(q27;q32)</b>
PC myeloma	<i>mum/irf4</i>	<b>t(6;14)(p25;q13)</b>

# Other dsDNA Breaks in B cells

Receptor editing

dsDNA breaks also must occur when a B cell switches from  $\kappa$  to  $\lambda$  light chain or vice versa

# Why Do Other dsDNA Breaks Occur at Specific Sites?

Unknown

Breakpoints usually occur within the introns; in some genes the breakpoints appear to cluster in certain regions of gene

Theories, none proven:

- Homologous recombination between Alu elements

- Cleavage at purine/pyrimidine repeat sequences

- Topoisomerase II subunit errors during DNA replication

- Non-homologous DNA repair during replication

Must create fusion gene with growth advantage

# How Do We Determine if Monoclonal Population with Translocation?

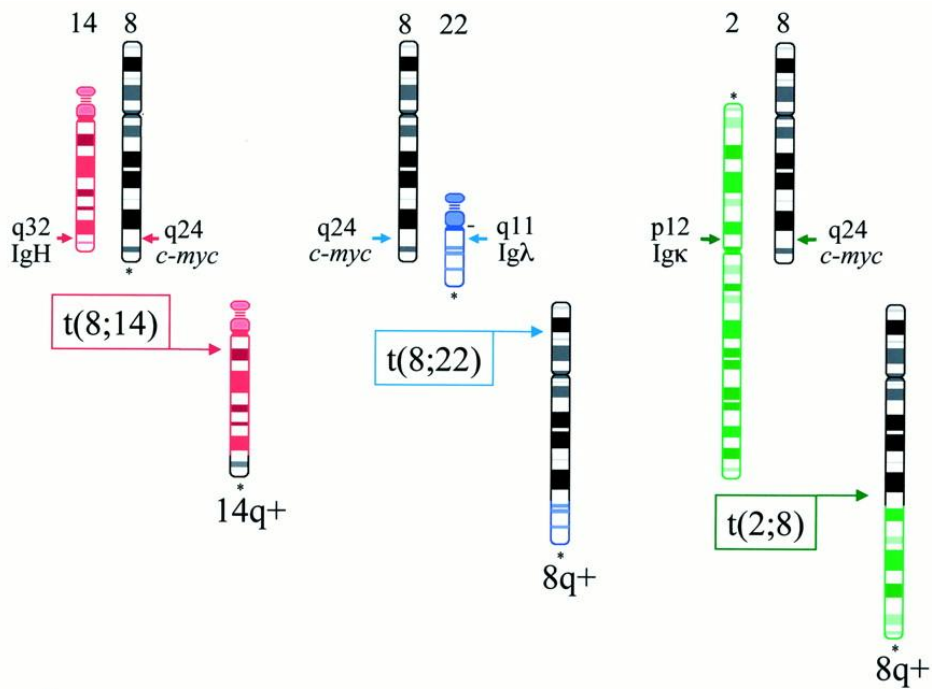
Cytogenetics

FISH - fluorescence in situ hybridization

RFLP-SB – restriction fragment length polymorphism-  
Southern blot

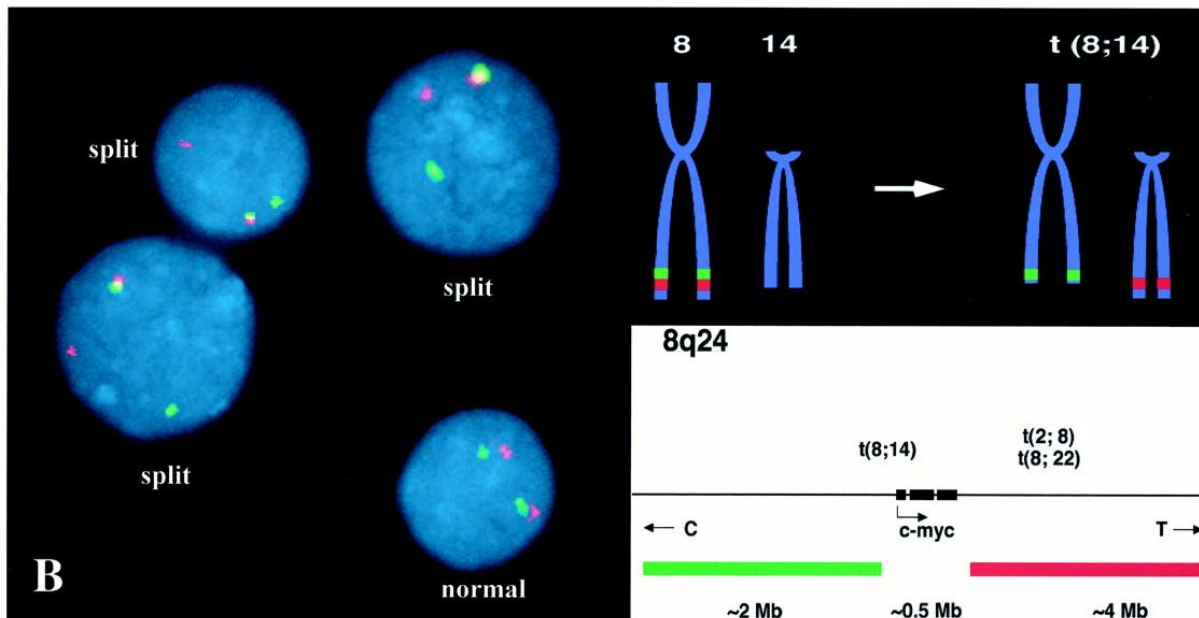
PCR or RT-PCR – need primers and probes

- PCR with primers straddling CDR3; run the product on:
  - polyacrylamide gel - EtBr stained - look for single strong band
  - ABI 310 capillary gel electrophoresis - fluorescent marker -  
look for large peak



A

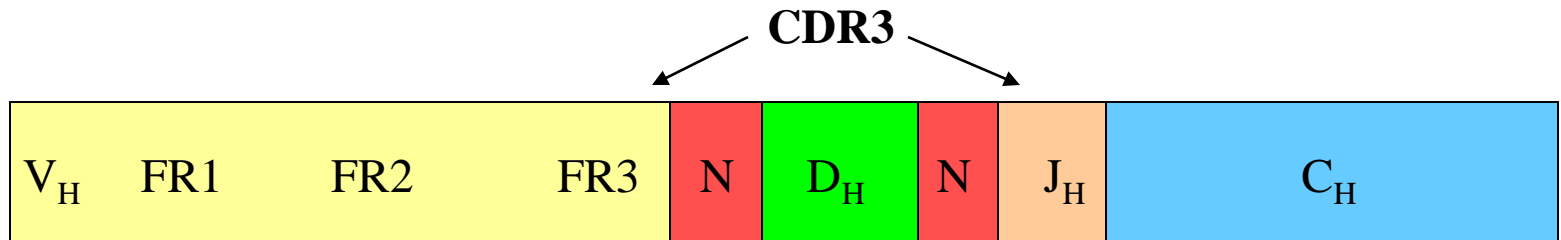
## FISH of *c-myc*/Ig translocations in Burkitt's



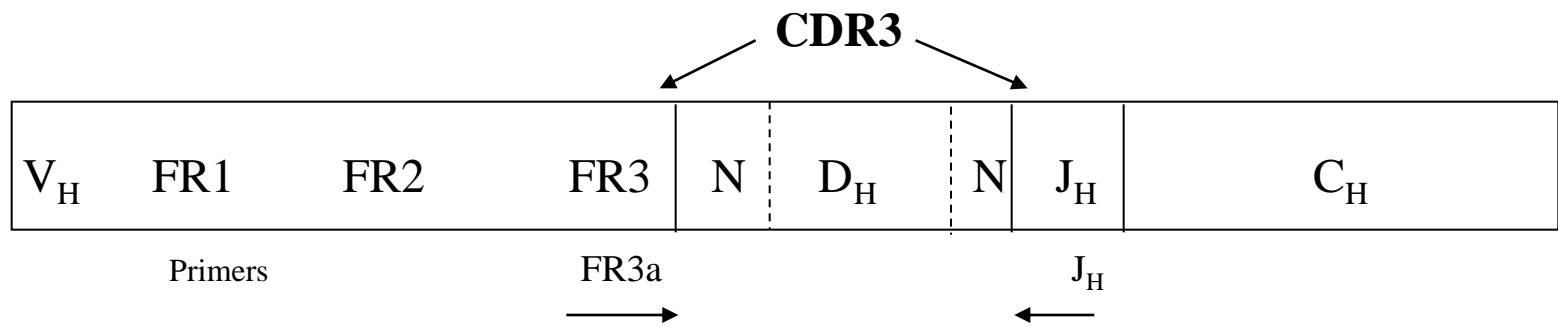
B

Hecht, J. L. et al. *J Clin Oncol*; 18:3707-3721 2000

# PCR of IgH Gene $\pm$ Translocation

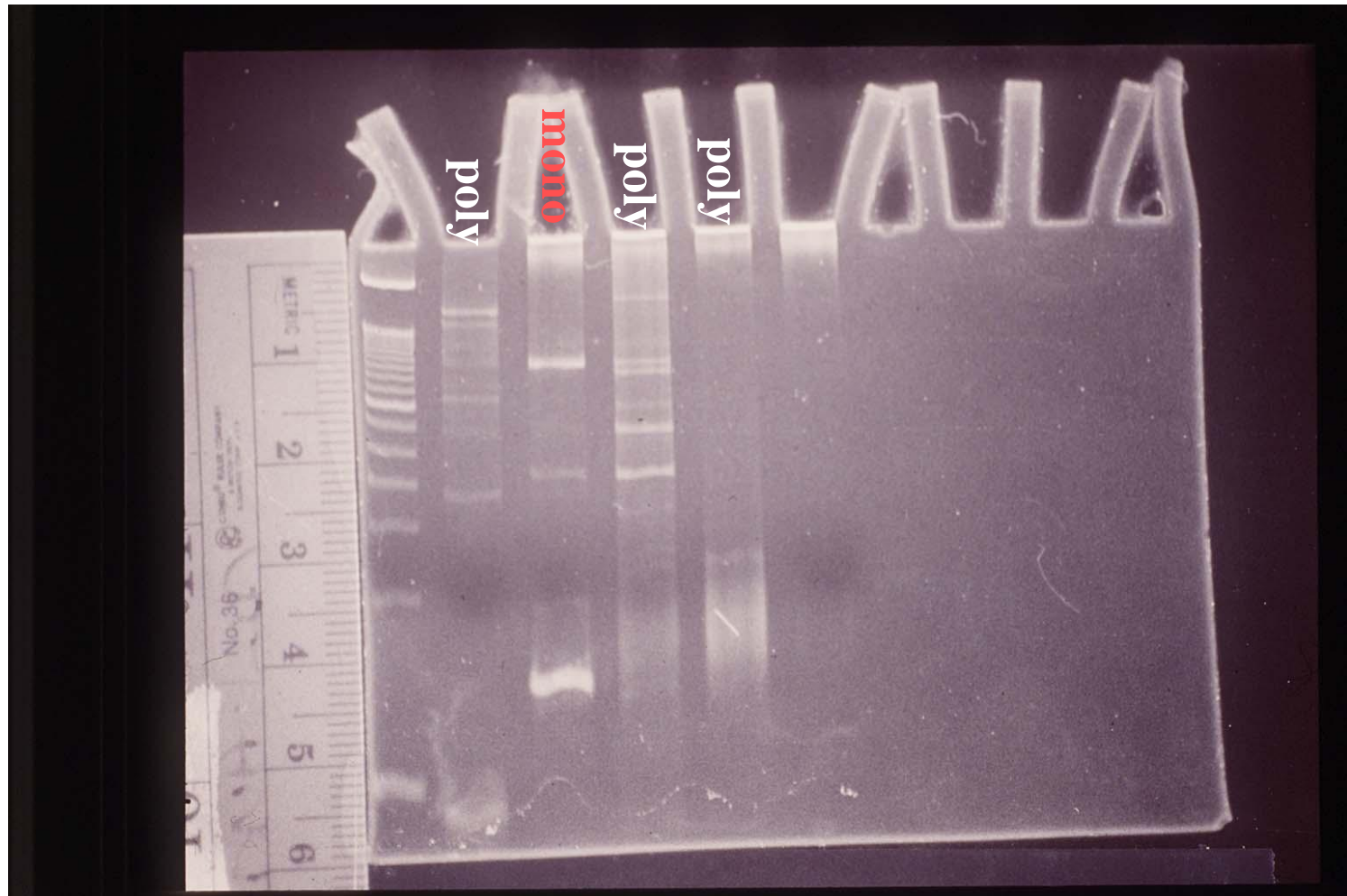


- BUT, point of cleavage is flexible
- AND, a few nucleotides in the coding regions may get trimmed during fusion
- ALSO, a few nucleotides ( $\leq 15$  N) may be added during recombination between

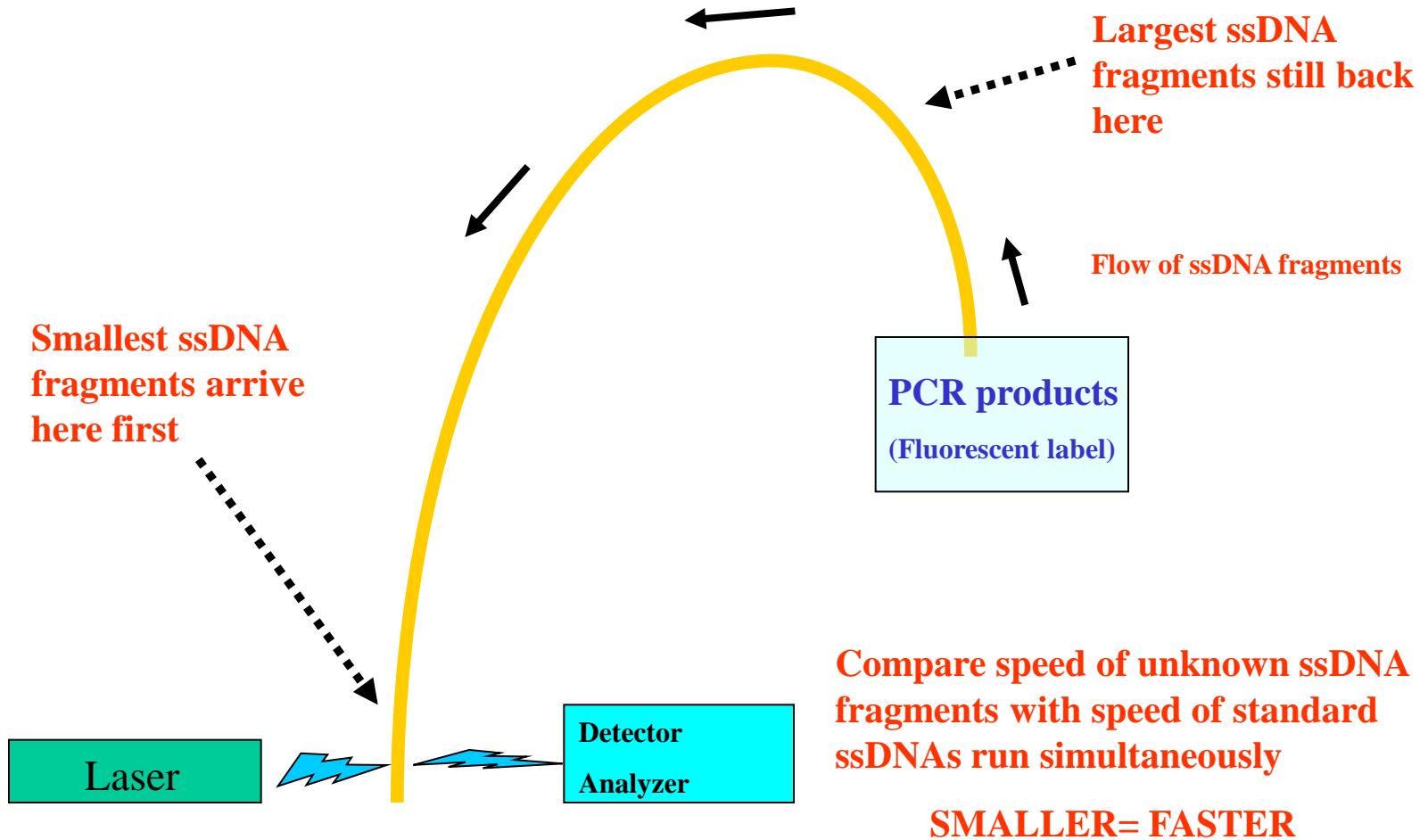




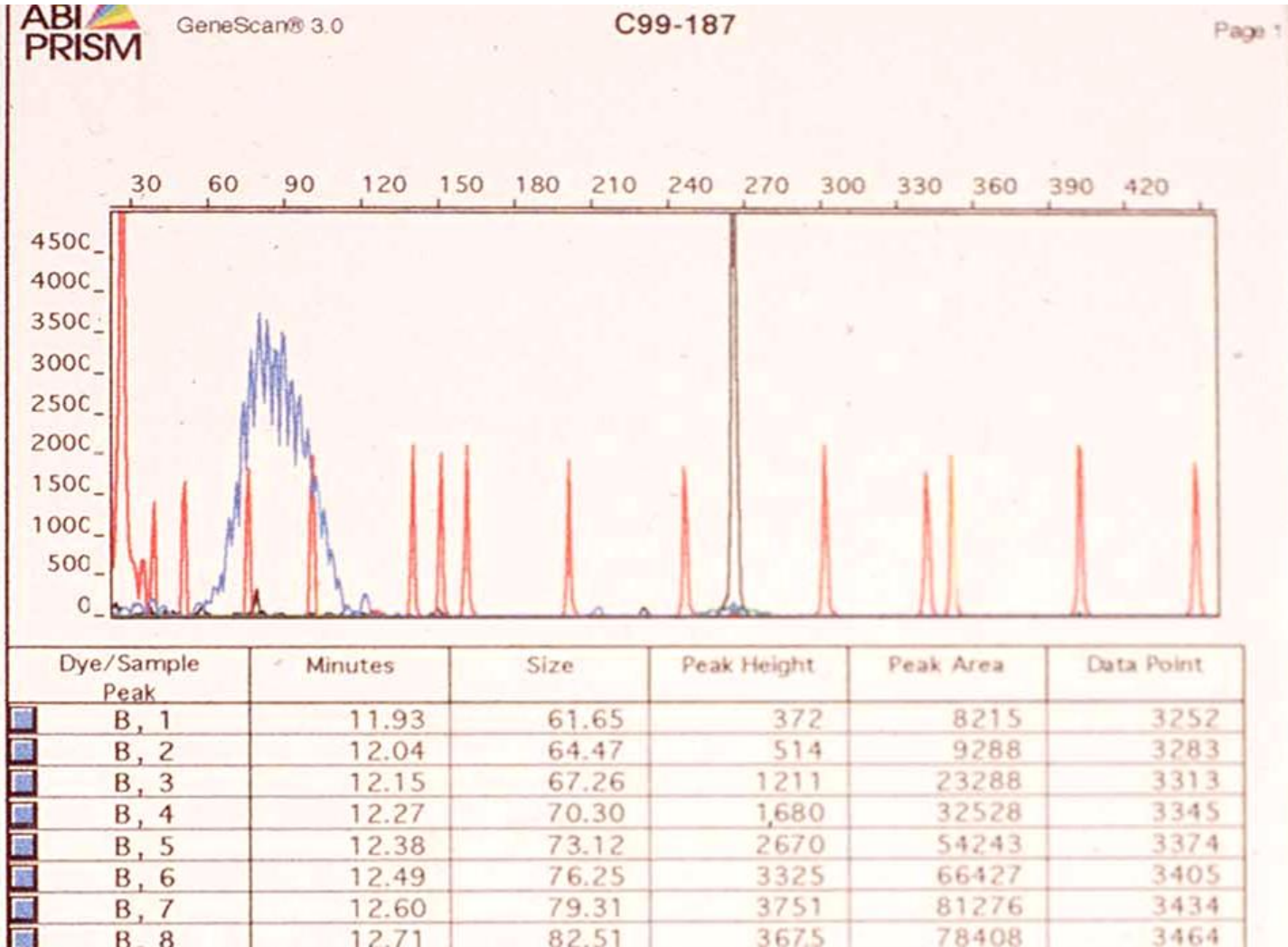
# PAGE Gel of PCR Products after amplification of FR3 in the IgH Gene



# Capillary Gel Electrophoresis

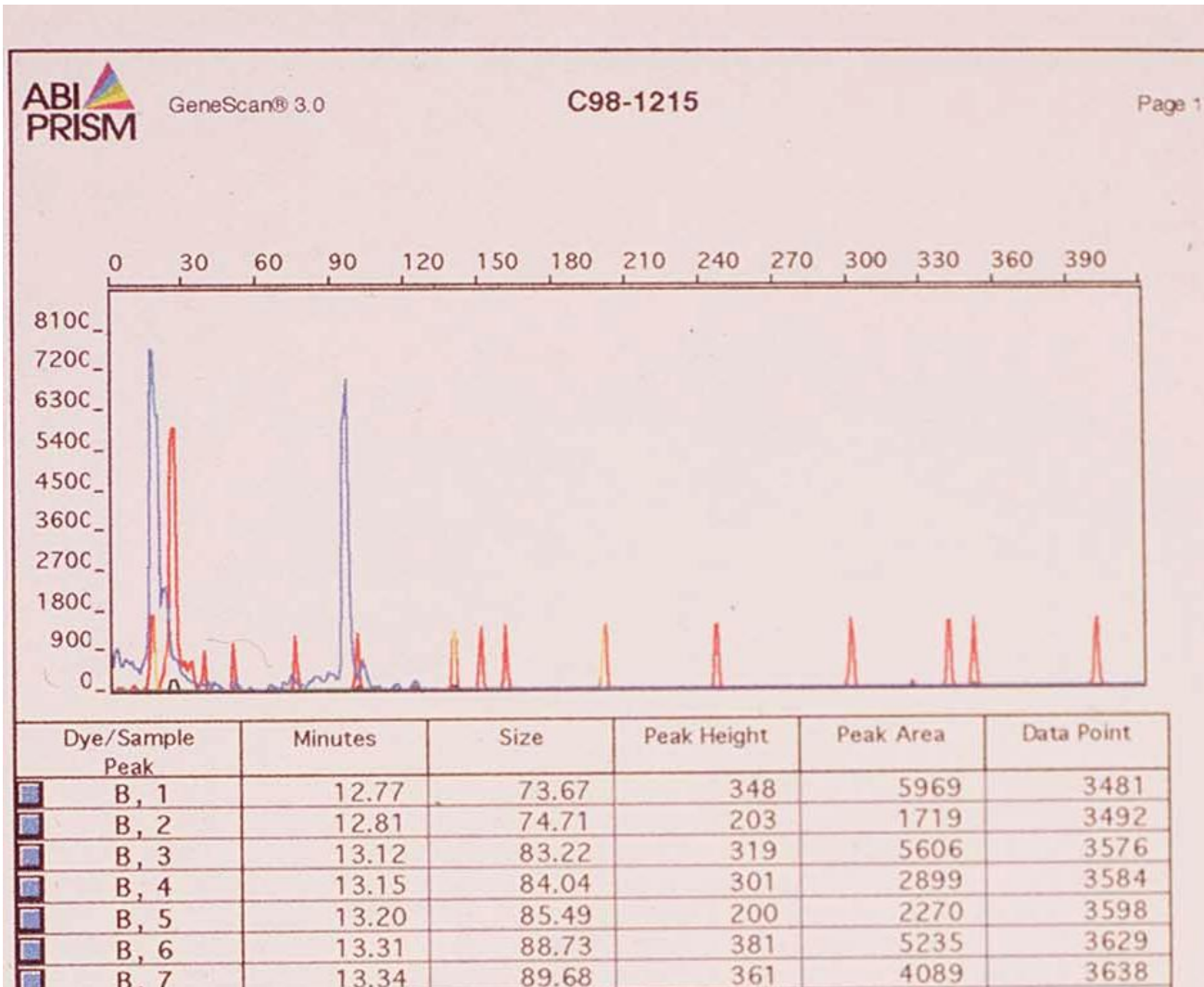


# Capillary gel electrophoresis of PCR products after amplification of FR3 in the IgH gene



**Polyclonal**

# Capillary gel electrophoresis of PCR products after amplification of FR3 in the IgH gene



**Monoclonal**

# Follicular Lymphoma

**t(14;18)(q32;q21)** in 90% of cases

Translocates the *bcl-2* gene on 18 next to enhancer and promoter elements of the IgH gene on chr 14

Results in the over-expression of the Bcl-2 protein, which has anti-apoptotic function

Necessary but not sufficient for tumor formation

Major breakpoint cluster region (MBR) – 60% of cases

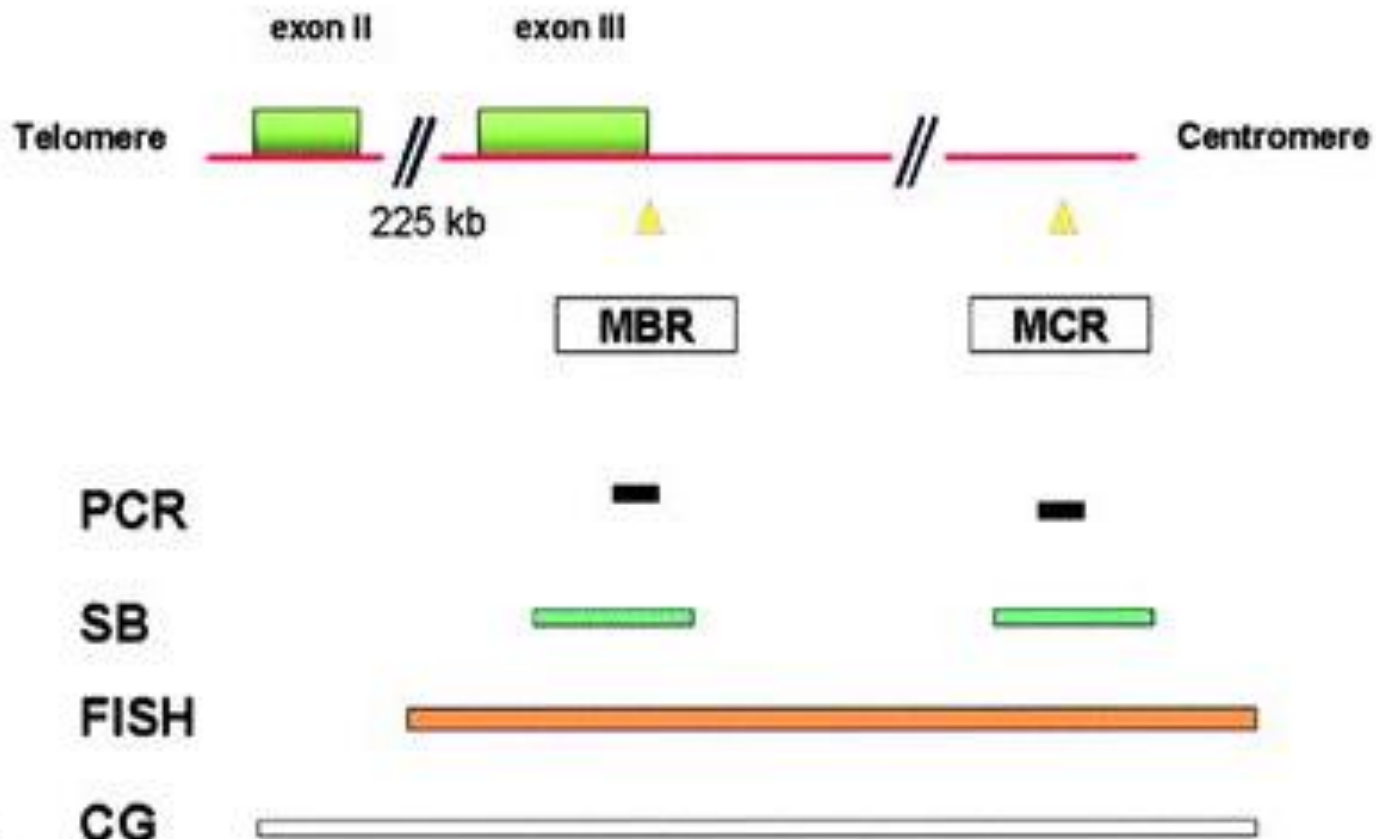
Minor breakpoint cluster region (MCR) – 15% of cases

Intermediate breakpoint cluster region – small % of cases

Detect by cytogenetics, FISH, PCR, quant PCR

# Follicular Lymphoma

← Chromosome band 18q21 →



# CLL and SLL

Translocations in only 5% of cases

Most have 13q14 deletion, trisomy 12, or 11q deletion

Translocations:

- t(2;18)(p12;q21)** Igκ gene translocated onto *bcl-2* gene

- t(18;22)(q21;q11)** Igλ gene translocated onto *bcl-2* gene

- t(14;19)(q32;q13)** *bcl-3* gene translocated onto IgH gene

- result in Bcl-2 or Bcl-3 overexpression

The t(14;19) seen in young age, poorer Px

Detection by cytogenetics, RFLP-Southern blot, PCR

# Mantle Cell

**t(11;14)(q13;q32)** in nearly all cases

Translocates the cell cycle regulator gene known as *cyclin D1/ccnd-1/bcl-1/prad1* of chr 11 onto the enhancer of the IgH gene of chr 14 near the J region

Results in cyclin D1 overexpression

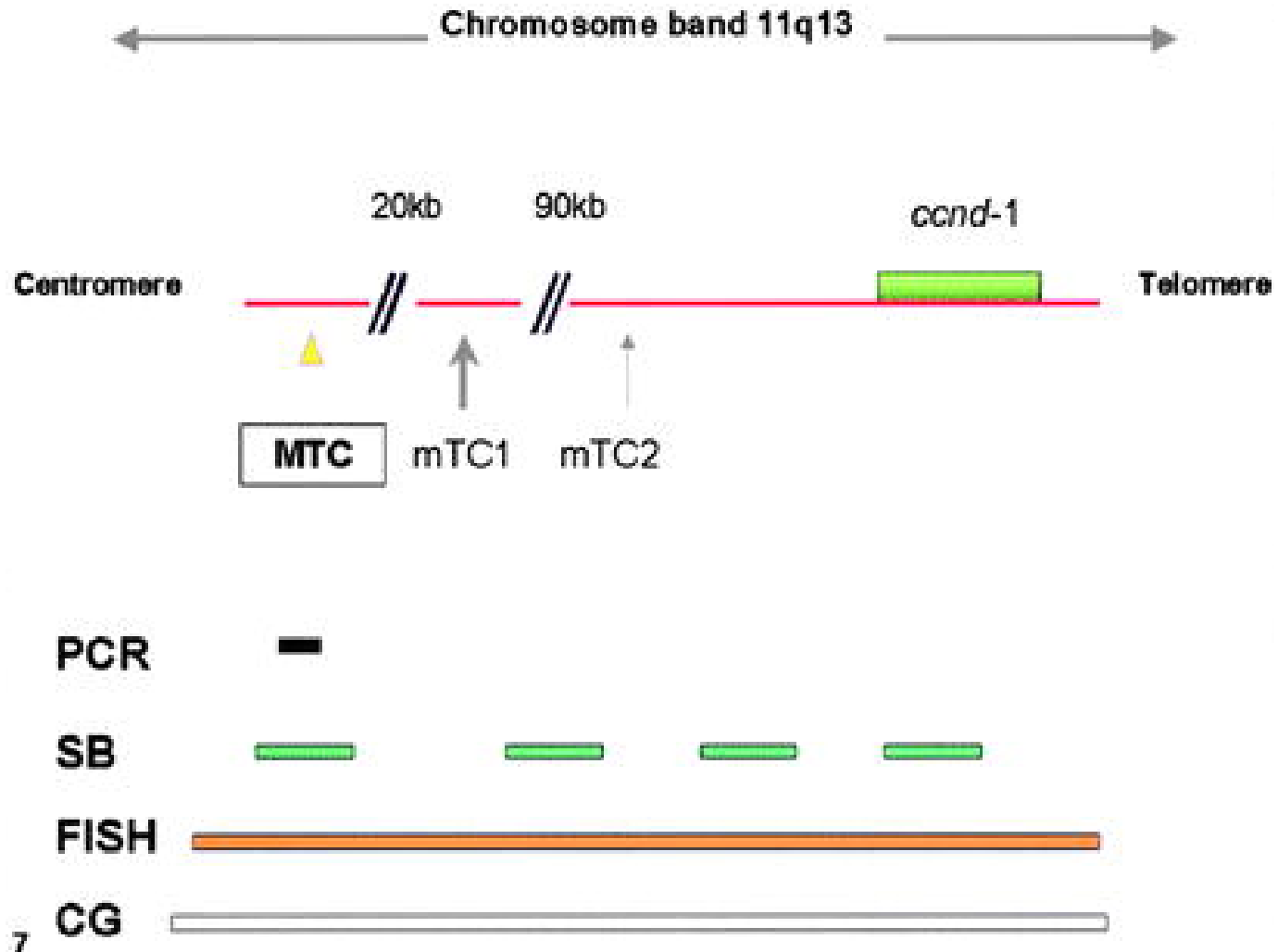
Major translocation cluster region in 11q13 - 50% of cases

Detection by cytogenetics, FISH, RFLP-SB, PCR, and quantitative RT-PCR

Immunohistochemistry – not great sensitivity



# Mantle Cell Lymphoma



# MALT Lymphoma

Multiple translocations described

**t(11;18)(q21;q21)** 35% of MALTs (most common one)

*api2* gene on chr 11 translocates to *malt1* gene on chr18

*api*= inhibitor of apoptosis

BIR= baculovirus inhibitor of apoptosis repeat CARD= caspase recruitment domain RING= really interesting new gene

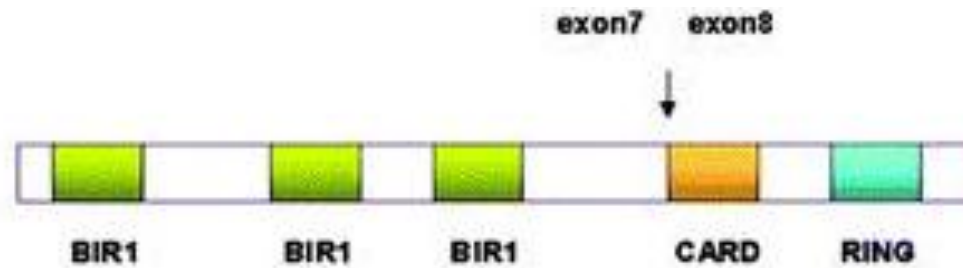
*malt1* functions in signaling pathway; structure is similar to Ig receptor

*malt1* has 4 main breakpoint sites in t(11;18)(q21;q21)

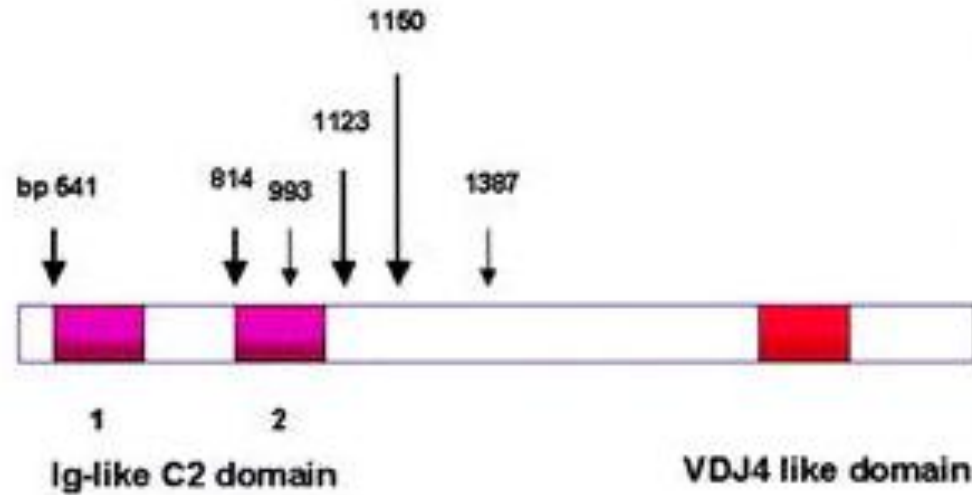
*malt1-api2* fusion leads to increased apoptosis inhibition, and NF-*k*B probably involved

# Malt Lymphoma

*api2*



*malt1*



# MALT Lymphoma

**t(11;18)(q21;q21)** 35% of MALTs (most common one)  
*malt1* function is unknown; probably involved in BCR signaling  
*malt1* has 4 main breakpoint sites in t(11;18)(q21;q21)

The *malt1-api2* translocation results in fusion protein, which functions to increase apoptosis inhibition and probably activate transcription factor NF- $\kappa$ B

## **Prognosis:**

- t(11;18) = usually not *H pylori* driven, no response to Abx  
= rare for high grade BCL to arise in these MALTs
- non t(11;18) = usually *H pylori* driven, do respond to Abx  
= transform to high grade BCL more often

# MALT Lymphoma

**t(14;18)(q32;q21)** 20% of MALTs

*malt1* gene on chr 18 translocated onto IgH gene on chr 14

Results in *malt1* overexpression

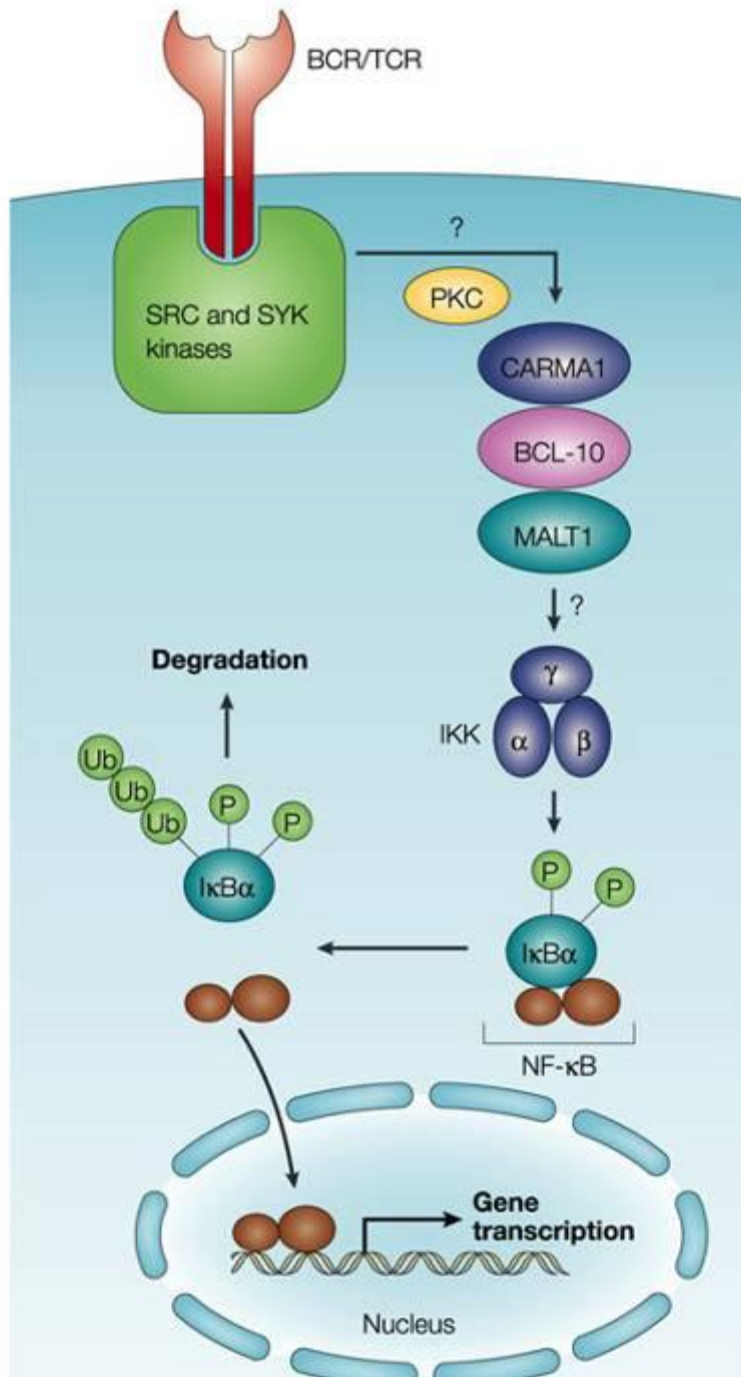
**t(1;14)(p22;q32)** <5%, new

*bcl-10* on chr 1 translocated onto IgH on chr 14

Results in *bcl-10* overexpression due to IgH enhancer

Normal Bcl-10 protein is a signaling protein, activates NF-*k*B, which promotes cell activation/replication

*bcl-10*-IgH fusion disrupts Bcl-10 protein function, may cause Bcl-10 deregulation→NF-*k*B activation, leading to cell proliferation?



**Malt1** and **Bcl-10** are thought to be part of signaling pathway for **NF- $\kappa$ B**

# MALT Lymphoma

**t(1;2)(q22;p12)** rare, new

Translocation of *bcl-10* gene on chr 1 onto Igκ gene on chr 2

Detection of all four translocations mainly by cytogenetics

- IHC to detect Bcl-10:

- normal B cells = Bcl-10 in cytoplasm

- MALT lymphoma = Bcl-10 in nucleus in t(1;14) and t(11;18) translocations

# Diffuse Large B-cell Lymphoma

Over 25 translocations reported, 40% of which involve *bcl-6*  
30% have the **t(14;18)(q32;q21)** seen in FL, prob transformed FL

*bcl-6* translocations seen with IgH, Igκ, Igλ, hsp90, eif4A11, et al  
*bcl-6* translocations and mutations also seen in MALT, FL, BL, CLL  
In fact, *bcl-6* alterations are the most common genetic defect in BCLs

Normal *bcl-6* is transcription repressor that regulates germinal center  
B-cell formation and development, antibody-affinity maturation,  
T<sub>H</sub> cell responses, plasma cell maturation



# Diffuse Large B-cell Lymphoma

In DLBCL, *bcl-6* gets translocated onto the promoter regions of the other genes, but the *bcl-6* coding region remains intact

Felt that *bcl-6* translocation onto other gene's promoter causes dysregulation of *bcl-6* and this leads to lymphoma

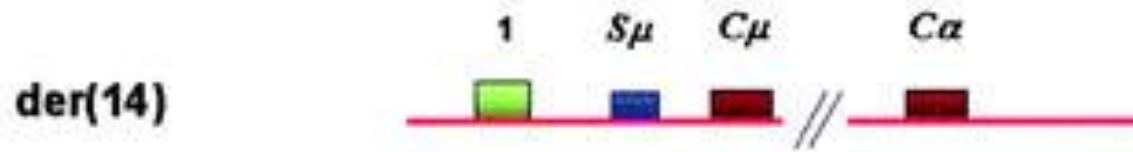
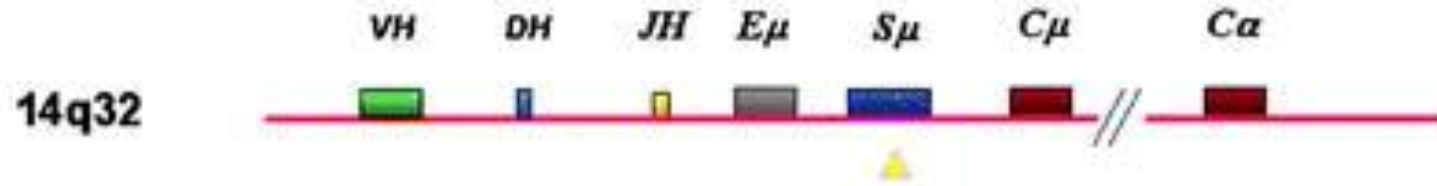
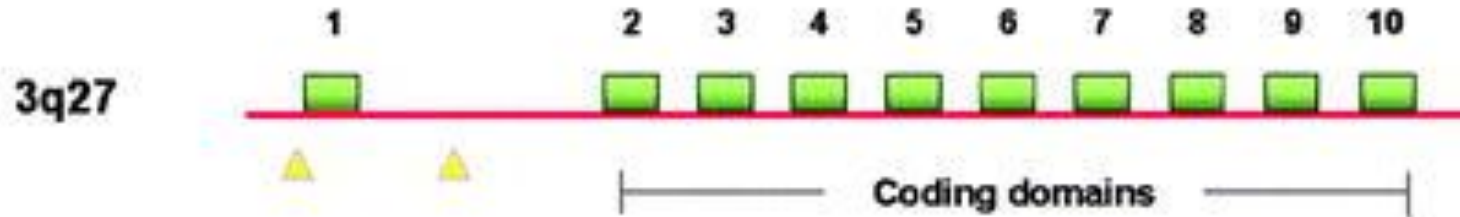
About 20% of DLBCL are Ig gene - *bcl-6* gene translocations:

- Ig gene translocations have better prognosis
- Non-Ig translocations have worse prognosis

Detection – RFLP-SB, FISH; not PCR (need too many primers)

# Diffuse Large B-cell Lymphoma

*bcl-6*



# Translocations in DLBCL

Translocation	Involved Genes
t(14;18)(q32;q21)	<i>IgH</i> and <i>bcl-2</i>
t(3;4)(q27;p13)	<i>bcl-6</i> and <i>rhoh/ttf</i>
t(3;6)(p27;p21.3)	<i>bcl-6</i> and <i>H4</i> histone
t(3;6)(p27;p21.2)	<i>bcl-6</i> and <i>pim-1</i>
t(3;7)(p27;p12)	<i>bcl-6</i> and <i>ikaros</i>
t(3;11)(p27;q23.1)	<i>bcl-6</i> and <i>bob/obf1</i>
t(3;3)(p27;q29)	<i>bcl-6</i> and <i>tfr</i>
t(3;13)(q27;q14)	<i>bcl-6</i> and <i>1-plastin</i>
t(3;16)(p27;p11)	<i>bcl-6</i> and <i>Il-21r</i>
t(3;12)(p27;12q23-24.1)	<i>bcl-6</i> and $\alpha$ - <i>nac</i>
t(3;18)(p27;p11.2)	<i>bcl-6</i> and <i>EIF4A1</i>
t(3;14)(p27;q32)	<i>bcl-6</i> and <i>hsp89<math>\alpha</math></i>
t(3;6)(p27;p12)	<i>bcl-6</i> and <i>hsp90<math>\beta</math></i>
t(3;16)(p27;p13)	<i>bcl-6</i> and <i>cllta</i>
t(3;14)(p27;q32)	<i>bcl-6</i> and <i>IgH</i>
t(3;2)(p27;p12)	<i>bcl-6</i> and <i>Ig<math>\kappa</math></i>
t(3;22)(p27;q11)	<i>bcl-6</i> and <i>Ig<math>\lambda</math></i>
t(14;15)(q32;q11-13)	<i>IgH</i> and <i>bcl-8</i>
t(1;22)(q22;q11)	<i>fcgRIIb</i> and <i>Ig<math>\lambda</math></i>
t(1;14)(q21;q32)	<i>muc-1</i> and <i>IgH</i>
t(10;14)(q24;q32)	<i>nf-<math>\kappa</math>2</i> and <i>IgH</i>
t(1;13)(p32;q14)	—
t(1;7)(q21;q22)	—
der(6)t(6;8)(q11;q11)	—
t(5;16)(?;q11-q12)	—
t(19;22)(q13;q11-q13)	—

# Burkitt's Lymphoma

**t(8;14)(q24;q32)** 85% of all cases most common type

*c-myc* gene on chr 8 translocates onto IgH gene on chr 14

Breakpoints on 14q and 8q vary, endemic vs sporadic BL

sporadic = 5' end or in *c-myc* → C or S region of IgH gene

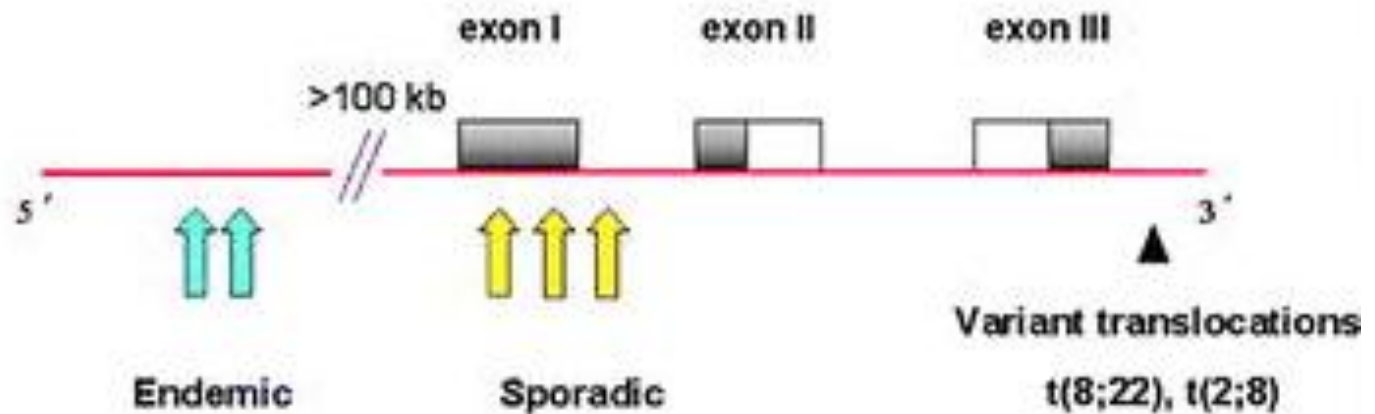
endemic = far 5' end of *c-myc* → J region of IgH gene

**t(2;8)(p12;q24)** *c-myc* gene translocated onto Igκ gene

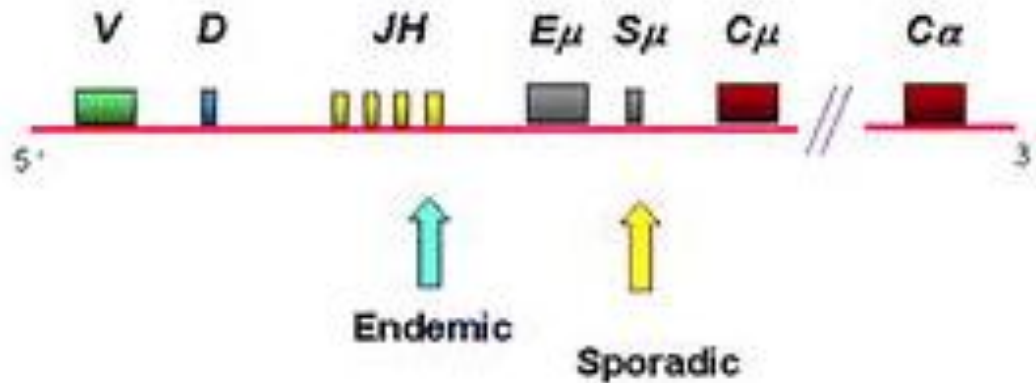
**t(8;22)(q24;q11)** *c-myc* gene translocated onto Igλ gene

# Burkitt's Lymphoma

Germline *c-myc* (8q24)



Germline *IgH* (14q32)



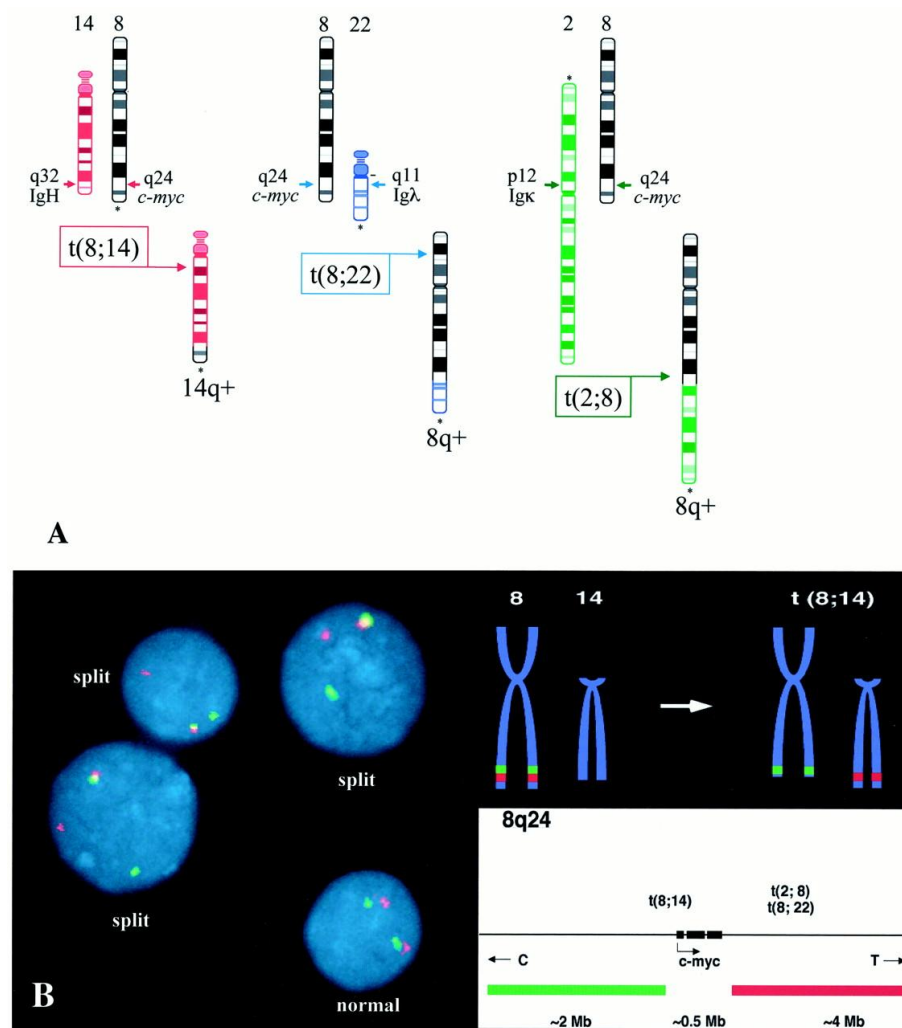
# Burkitt's Lymphoma

Pathogenesis- in all 3 the *c-myc* gene is overexpressed due to translocation next to Ig gene enhancers

Normal *c-myc* regulates cell proliferation, differentiation, apoptosis, cell cycle progression

Detection – cytogenetics, FISH, RFLP-SB on some cases, but not PCR (too many primers needed)

# Detection of *c-myc*/Ig chromosomal translocations in Burkitt's



Hecht, J. L. et al. J Clin Oncol; 18:3707-3721 2000

# Anaplastic Large Cell Lymphoma

**t(2;5)(p23;q35)** most common, in 50-70% of cases

Nucleophosmin (*npm*) gene of chr 5 translocated onto anaplastic lymphoma kinase (*alk*) gene of chr 2

Normal NPM = rRNA processing; nuclear transport protein

Normal ALK = membrane protein with intracytoplasmic tyrosine kinase; only expressed in CNS and PNS

ALK expression in hematopoietic cells is abnormal



# Anaplastic Large Cell Lymphoma

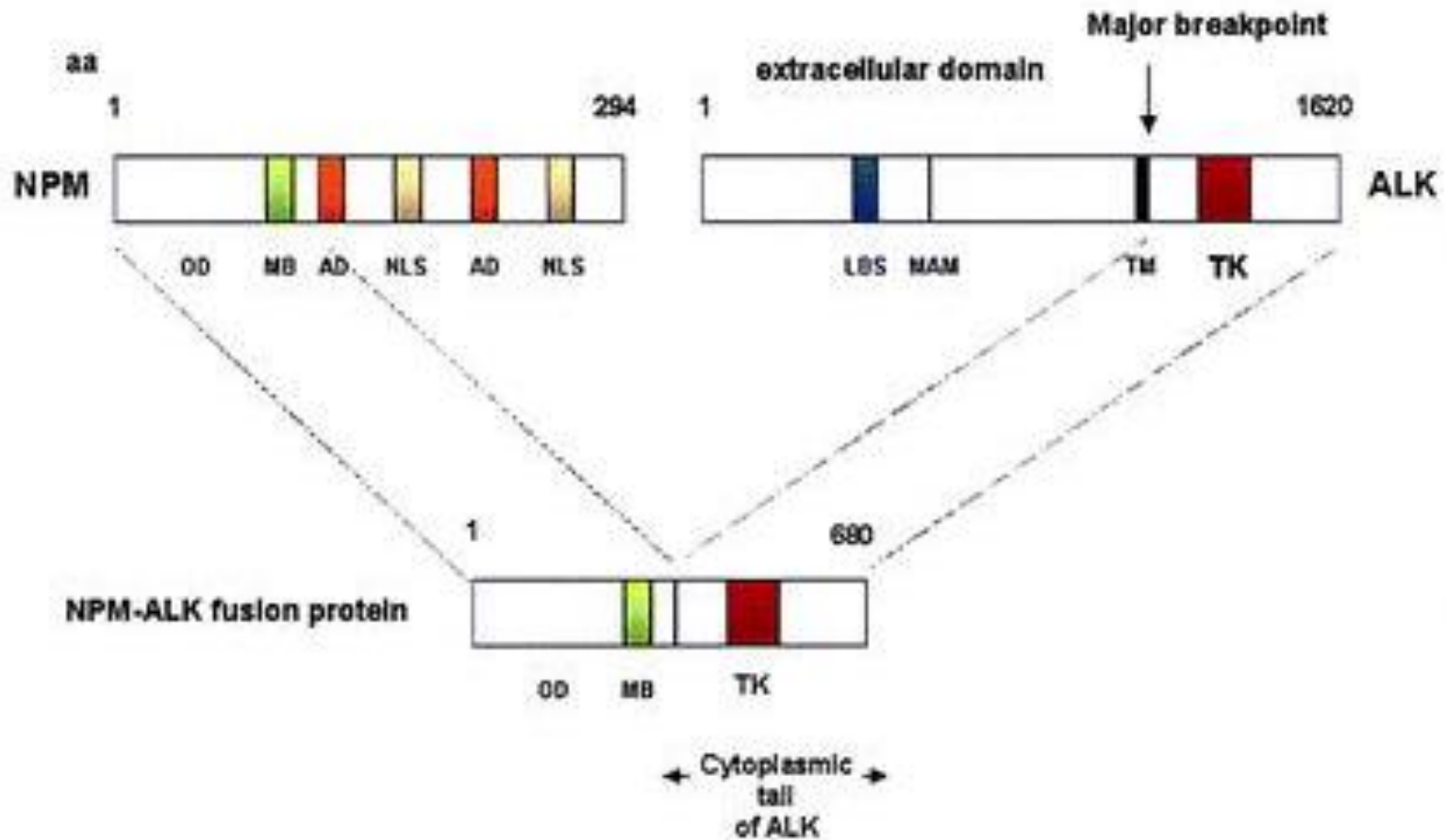
Fusion of *npm* and *alk* genes leads to encoded NPM-ALK fusion protein expression; *npm* promoter drives NPM-ALK transcription

NPM-ALK fusion protein is thus constantly expressed and ALK's tyrosine kinase activity turned on all the time

ALK tyrosine kinase stimulates downstream mitogen signaling and constant proliferation signals to the B-cell

Detection – cytogenetics, FISH, IHC (usually both cytoplasmic and nuclear stain), but not PCR

# Anaplastic Large Cell Lymphoma



# Plasma Cell Myeloma

Several translocations reported and most involve IgH on 14q32

**t(11;14)(q13;q32)** 30% of cases

*ccnd-1* (cyclin D1) gene on chr 11 transloc onto IgH on chr 14

*ccnd-1* breakpoints not the same as those in mantle cell

IgH gene breakpoints are in the S (switch) region

**t(4;14)(p16;q32)** 25% of cases

*fgfr3* on chr 4 translocated onto IgH of chr 14

*fgfr3* (fibroblast growth factor receptor 3) gene is dysregulated

# Plasma Cell Myeloma

**t(6;14)(p25;q32)** 20% of cases

*mum/irf4* gene of chr 6 translocated onto IgH gene

*mum/irf4* (multiple myeloma gene 1/interferon regulatory factor 4) dysregulated due to IgH enhancer

**t(14;16)(q32;q23)** 10% of cases

*c-maf* gene on chr 16 translocated onto IgH gene

c-Maf (musculoaponeurotic fibrosarcoma) protein is a transcription factor that helps regulate expression of cyclin D1 and IL-4

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These translocations seen in MGUS and PC myeloma

They are likely initiating translocations, and further mutations needed for full blown myeloma

Myelomas also frequently have non-translocation chromosomal defects

Detection – FISH usually; cytogenetics; PCR works some of the time